Canvax Reagents, S.L.U.
Luis de Mercado Street, 19
Boecillo Technological Park
47151, Boecillo
Valladolid, Spain.

Tlf: +34 983 54 85 63
info@canvaxbiotech.com

www.canvaxbiotech.com

DATA SHEET

Version: 04
Revision date: 17/07/2025

1. Identification

Product name **Clean-Easy PCR & Gel Purification Kit**

Cat. No **AN0091 (100 reactions)**
Cat. No **AN0091-XL (250 reactions)**

2. Description

Clean-Easy PCR & Gel Purification Kit, developed as a 2-in-1, provides a rapid and efficient method to purify DNA and remove contaminants from reaction mixtures (e.g. PCR, digestion or labelling reactions) as well as extraction of DNA from both TAE and TBE agarose gels.
The DNA is bound to silica membrane in presence of chaotropic salts, washed and then eluted in a separate tube. The purified DNA is ready to be used in all demanding molecular biology applications, including restriction digestion, ligation, sequencing, transfection into mammalian cells and in vitro transcription.

3. Kit Components

Item	Quantity	
	AN0091 (100 rxn)	AN0091-XL (250 rxn)
Clean-Easy minispin columns	100	250
Collection tubes (2 mL)	100	250
Buffer QG	2 x 60 ml	5 x 60 ml
PE Buffer*	25 ml	40 ml
EB Buffer	10 ml	20 ml

* Ethanol (96%-100%) [not included] must be added prior to use as indicated on the label. After ethanol has been added, mark the bottle to indicate that this step has been completed.

4. Features

- **Dual performance kit** for both PCR product cleanup and DNA purification from agarose gels
- **Simple** and **Just a few minutes** procedure
- DNA purified **Ready to use** in all molecular biology procedures
- **Wide spectrum of size** fragments could be purified. From 75bp to a maximum of 50kb.
- **Efficient DNA recovery.** Yields of up to 90% depending on fragment size
- **High binding capacity.** Each column handles up to 10 µg of DNA

5. Storage specifications

Clean-Easy PCR & Gel Purification Kit should be stored at room temperature (15–25˚C) for up to 12 months without any reduction in performance.

6. Applications

- Fast purification of DNA from agarose gels.
- Fast purification of products from PCR amplification reactions.

Canvax Reagents, S.L.U.
Luis de Mercado Street, 19
Boecillo Technological Park
47151, Boecillo
Valladolid, Spain.

Tlf: +34 983 54 85 63
info@canvaxbiotech.com

www.canvaxbiotech.com

DATA SHEET

Version: 04
Revision date: 17/07/2025

7. Further information

Product Use Limitations	This product is developed, designed, and sold exclusively only for research purposes use. The product was not tested for use in diagnostics or for drug development, nor is it suitable for administration to humans or animals.
Safety Information	When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, please consult the appropriate material safety data sheets (MSDS). These are available online in convenient and compact PDF format at www.canvaxbio.com where you can find, view, and print the MSDS for each CANVAX kit.

1. SAMPLE PREPARATION

PCR Clean Up

1. Add 5 volumes of **Buffer QG** to one volume of PCR solution and mix thoroughly by pipette.

Gel Extraction

1. Using a clean, sharp razor blade or scalpel, excise the DNA ban from the agarose gel. Remove the extra agarose to reduce the size of gel slice. Place the gel slice in a 1.5 ml pre-weighted tube and weigh the gel slice (The maximum amount of gel slice per column is 400 mg).

2. Add 3 volumes of **Buffer QG** to 1 volume of gel. (For example, if the agarose gel slice is 100 mg, add 300 µl of Buffer QG)

*For gels containing more than 2% agarose, add 6 volumes de **Buffer QG** per mg of gel.*

3. Incubate at 50 ˚ C in a water bath for 10 min or until the gel slice has completely dissolved. During incubation at 50 °C, mix by vortexing or inverting the tubes every 1 minute. **Make sure the gel slice completely dissolved. For >2% gels, increase incubation time.**

Important! *For fragments <500 bp and >4 kb, add 1 volume of isopropanol to the sample and mix (For example, if the agarose gel slice is 100 mg, add 100 µl isopropanol)*

2. DNA BINDING

1. Label the lid of a new spin column placed in a 2 ml collection tube. Carefully apply the mix from step 1 (1 *Sample preparation*) to the spin column and Centrifuge at 13000 rpm for 1 minute.

2. Place the spin column in a new 2 ml collection tube and discard the collection tube containing the filtrate.

3. WASH

1. Add 700 µl of **buffer PE** for Wash to the minispin column and centrifuge at 13000 rpm for 1 minute.

Remember! Before using it for the first time, add ethanol (96–100%) to the PE Buffer as indicated on the bottle.

2. Discard the flow-through and centrifuge at 13000 rpm for 1 minute. *This step is essential for removing traces of PE buffer.*

3. Transfer the minispin column into a new, labelled 1.5 ml microcentrifuge tube.

4. DNA ELUTION

1. Carefully open the minispin column and pipet 30 µl Buffer EB or H2O (pH=7.0-8.5) directly onto the membrane. Close the cap and incubate for 1 min at room temperature, then centrifuge at 13000 rpm for 1 min to elute DNA. *To increase the DNA yield you can warm the buffer EB/H2O to 65 ˚C before adding to the column*.